

Minireview: From Anorexia to Obesity—The Yin and Yang of Body Weight Control

JEFFREY M. ZIGMAN AND JOEL K. ELMQUIST

Department of Medicine and Division of Endocrinology (J.M.Z., J.K.E.), and Department of Neurology (J.K.E.), Beth Israel Deaconess Medical Center, and Program in Neuroscience (J.K.E.), Harvard Medical School, Boston, Massachusetts 02215

Over the past decade, there has been a tremendous increase in the understanding of the molecular and neural mechanisms that control food intake and body weight. Yet eating disorders and cachexia are still common, and obesity cases are rising at alarming rates. Thus, despite recent progress, an increased understanding of the molecular and neural substrates that control body weight homeostasis is a major public health goal. In this review, we discuss the mechanisms by which metabolic signals interact with key behavioral, neuroendocrine, and au-

tonomic regulatory regions of the central nervous system. Additionally, we offer a model in which hormones such as leptin and ghrelin interact with similar central nervous system circuits and engage them in such a way as to maintain an appropriate and tight regulation of body weight and food intake. Our model predicts that overstimulation or understimulation of these central pathways can result in obesity, anorexia, or cachexia. (*Endocrinology* 144: 3749–3756, 2003)

OBESITY, EATING DISORDERS, and cachexia endanger the lives of millions of people worldwide. Fortunately, during the last decade, there has been rapid and substantial progress toward uncovering the molecular and neural mechanisms by which these extremes of energy balance develop. Central to this research has been the identification and characterization of certain peripheral metabolic signals, including leptin and ghrelin, which serve as fundamental indices of energy sufficiency.

Several recent reviews have discussed in detail many components of neuroendocrine control of body weight (1–6). In this review, we will outline examples of central mechanisms that underlie the adaptive responses that occur in states of energy abundance or insufficiency. We will focus much of our attention on the hormones leptin and ghrelin, both of which act as crucial signals at either end of the energy spectrum. Clearly, other metabolic signals contribute to body weight homeostasis but can only be briefly mentioned (5, 7, 8). Finally, we will offer a model in which leptin and ghrelin interact with similar central nervous system (CNS) circuits, including several hypothalamic and brainstem nuclei, setting into motion an integrated, coordinated, and adaptive response to the particular state of energy balance and food availability. Our model predicts that overstimulation or understimulation of these pathways result in obesity, anorexia, or cachexia. Such responses have undoubtedly been opera-

tional throughout evolution but are now being increasingly elaborated in the world in which we now live.

Leptin: A Molecular Signal of Energy Abundance

A state of energy abundance is generated when food intake and nutrient absorption exceed total body energy expenditure (3, 6). This positive energy balance is represented by increases in several circulating factors, including glucose, leptin, glucagon-like peptide-1 (GLP-1), and peptide YY₃₋₃₆ (PYY₃₋₃₆) (1, 3, 6). Among these, leptin, which is secreted by white adipose tissue, is established as the prototypical hormone released normally in an environment of nutritional plenty. Many studies indicate that both leptin's mRNA levels and its circulating levels are elevated in most obese humans and also in rodents with diet-induced obesity (4, 9, 10). Although relatively little is understood regarding the actual molecular controls of leptin production and secretion, leptin levels in environments of energy abundance are likely influenced by insulin and glucocorticoids (11–14); see also accompanying review by Rajala and Scherer (15).

Following its release, leptin acts both in the brain and in peripheral tissues (16–25). In the CNS, leptin directly activates or inhibits neurons expressing the functional long leptin receptor (Ob-Rb) (22, 23). These receptors are located in several sites, including the hypothalamic arcuate nucleus (Arc), ventromedial hypothalamic nucleus (VMH), and dorsal medial hypothalamic nucleus (16–19). The leptin receptor is a type 1 cytokine receptor, which exerts its effects by activating the janus-kinase/signal transducer and activator of transcription-3 (STAT-3) pathway (26, 27). Leptin administration induces STAT-3 translocation, phosphorylation of phosphoinositide 3-kinase and the expression of several leptin-responsive genes, including suppressor of cytokine signaling-3 and *c-fos* in the hypothalamus and brainstem (20, 21, 28–36).

Abbreviations: AgRp, Agouti-related gene product; Arc, Arcuate nucleus; CART, cocaine- and amphetamine-regulated transcript; CNS, central nervous system; DVC, dorsal vagal complex; GHSR, GH secretagogue receptor; GLP-1, glucagon-like peptide-1; 5-HT₂CR, serotonin 2C receptor; LHA, lateral hypothalamic area; LPS, lipopolysaccharide; MCH, melanin-concentrating hormone; MC4R, melanocortin-4 receptor; NPY, neuropeptide Y; Ob-Rb, functional long leptin receptor; POMC, proopiomelanocortin; PVH, paraventricular hypothalamic nucleus; PYY₃₋₃₆, peptide YY₃₋₃₆; UCP-1, uncoupling protein-1; VMH, ventromedial hypothalamic nucleus.

Despite evidence clearly linking leptin to positive energy balance and to pathways that suppress ingestive behavior (see below), obesity often persists despite leptin elevations [see accompanying review by O'Rahilly *et al.* (37)]. This apparent leptin resistance can be viewed either as detrimental, given the morbidities associated with obesity, or as evolutionarily favorable, because it permits accumulation of further energy stores that potentially could be used during times of low environmental availability (9). Indeed, a fall in circulating leptin levels, as occurs during caloric restriction, is likely a more effective and critical signal to the CNS, promoting adaptation to a fasted state by increasing appetite, decreasing energy expenditure and modifying neuroendocrine function in a direction that favors survival (9).

Ghrelin: A Long-Sought Molecular Signal of Energy Insufficiency

Ghrelin is now established as an important indicator of energy insufficiency. Ghrelin is released mainly from endocrine cells of the stomach and gastrointestinal tract (38). It was originally identified in 1999 during a search for the endogenous ligand of the GH secretagogue receptor (GHSR; ghrelin receptor), which previously had been localized to some peripheral tissues and also to several CNS sites, including many of the same regions where Ob-Rb is found, such as the Arc and VMH (38–45). Human studies have found a preprandial rise and a postprandial decline in plasma ghrelin levels, suggesting that ghrelin plays a physiological role in hunger and meal initiation (46, 47). Fasting-associated elevations in ghrelin also occur in animals, both acutely and with chronic food deprivation (48, 49). Many other studies also clearly demonstrate that these elevations in ghrelin have a functional consequence aimed at reversing the state of energy deficit in which they arise. For example, anti-ghrelin IgG administration suppresses feeding in rodents, and both central and peripheral ghrelin administration strongly stimulate feeding, suppress energy expenditure, and lead to increased body weight (48, 50–53).

Adding further support to ghrelin's role as a signal of energy insufficiency are observations of altered ghrelin levels in human obesity and after weight loss. Initial studies revealed that fasting plasma ghrelin levels are negatively correlated with percentage of body fat and body weight (54). Following diet-induced weight loss, plasma ghrelin levels increase significantly, suggesting a mechanism for the rebound weight gain following dieting (47). Conversely, gastric bypass markedly suppresses ghrelin levels, which likely contributes to the weight-reducing effect of and the maintenance of the reduced weight associated with the procedure (47). Furthermore, in patients with Prader-Willi syndrome, high ghrelin levels are present and are thought to directly contribute to the voracious appetite, hyperphagia, and obesity that characterize this syndrome (55, 56).

Leptin and Ghrelin Activate Overlapping Pathways

As noted above, several hypothalamic cell groups express leptin and/or ghrelin receptors, and evidence is now accumulating to suggest that some of these nuclei, such as the Arc,

are critical sites of integration for leptin-responsive and ghrelin-activated pathways. Within the Arc, two distinct leptin- and ghrelin-responsive cell groups exist. The first is identified by the coexpression of POMC (proopiomelanocortin) and CART (cocaine- and amphetamine-regulated transcript) and is often referred to as an anorexigenic population (32). The second distinct population of neurons is orexigenic and coexpresses the peptides NPY (neuropeptide Y) and AgRP (agouti-related gene product) (3, 57–60). The leptin and ghrelin receptors are expressed in both Arc subpopulations (61, 62). Leptin activates POMC/CART neurons at the level of gene transcription and also by direct depolarization, presumably initiating the release of the two potent anorexigenic neuropeptides α -MSH (a melanocortin receptor agonist) and CART (21, 22, 63–66). In parallel, leptin directly inhibits NPY and AgRP transcription and hyperpolarizes the NPY/AgRP neurons, thus preventing the release of the potent orexigenic neuropeptides NPY and AgRP (the endogenous melanocortin receptor antagonist) (21, 23, 57, 66–68).

Consistent with ghrelin action in the Arc, ablation of the Arc with monosodium glutamate significantly blunts the ingestive behaviors normally stimulated by central delivery of ghrelin (69). In contrast to leptin, ghrelin activates arcuate NPY/AgRP neurons. This has been demonstrated by ghrelin- and/or GHSR agonist-stimulated *c-fos* induction and augmentation of NPY and AgRP transcription in NPY/AgRP neurons (50, 52, 70–73).

Evidence of an overlap of ghrelin- and leptin-responsive pathways in the Arc includes the ability of ghrelin to depolarize the majority of Arc neurons that are inhibited by leptin (74). Recent work by Cowley *et al.* (75) has demonstrated that ghrelin also directly depolarizes arcuate NPY/AgRP neurons. Additionally, administration of a GHSR agonist to obese, leptin-resistant *fa/fa* rats results in double the amount of arcuate *c-fos* induction normally seen (in lean animals), whereas conversely, chronic central infusion of leptin to normal, fasted animals suppresses this GHSR agonist-induced *c-fos* response (76). This suggests that lack of a functional leptin signaling pathway increases sensitivity to ghrelin. Furthermore, antibodies and antagonists of both NPY and AgRP abolish ghrelin-induced feeding, whereas NPY antagonists also block ghrelin's inhibitory effects on oxygen consumption (50, 51). Interestingly, administration of a GHSR agonist to NPY-deficient mice still stimulates food intake and body weight increases; these responses are blocked upon exposure to a melanocortin receptor agonist (77).

Yet another level of control of these circuits exists, in addition to the above-described direct actions of leptin and ghrelin on neurons in the Arc. Specifically, NPY/AgRP neurons provide a local (collateral) input to their neighboring arcuate POMC/CART cells (7, 22, 75). These NPY collaterals release the inhibitory neurotransmitter γ -aminobutyric acid, resulting in a tonic inhibition of the POMC/CART neurons (78). Importantly, leptin and ghrelin (as well as other signals, such as PYY₃₋₃₆) modulate this local circuit. For example as noted above, leptin directly depolarizes POMC neurons while simultaneously hyperpolarizing NPY/AgRP neurons. The inhibition of NPY/AgRP neurons results in a loss of inhibitory (γ -aminobutyric acidergic) input to POMC neu-

rons and a net increase in POMC activity (disinhibition). In contrast, ghrelin directly depolarizes the NPY/AgRP neurons and simultaneously increases inhibitory input to the POMC cells (75). This local circuit increases the ability to fine tune responses to changing energy availability.

Taken together, these observations support the hypothesis that inverse changes in leptin and ghrelin levels likely are critical to the maintenance of energy homeostasis, and it is intriguing to speculate on the effects of manipulating specific components of the abovementioned leptin- and ghrelin-responsive circuit. For example, specific deletion of either leptin receptors in POMC neurons or ghrelin receptors in NPY/AgRP arcuate neurons would be predicted to have effects on food intake and body weight. Moreover, manipulations in the NPY/AgRP neurons also would be predicted to affect both the tonic firing of POMC neurons and the indirect responses following changes in leptin or ghrelin levels. Obviously, these and other questions need to be addressed directly and need to be assessed in mice with cell-specific genetic deletions or lesions in various components of the aforementioned circuits. Undoubtedly, these types of studies represent where the field will move in the ensuing years. Despite the lack of definitive data currently, this local circuitry within the Arc is likely an important modifier of the responses to changes in key metabolic hormones including leptin, ghrelin, insulin, and PYY₃₋₃₆ (7, 22, 23, 75, 79).

Direct Leptin and Ghrelin Action in the Brainstem

Although we have concentrated on the actions of metabolic signals in the hypothalamus, a growing body of evidence suggests that direct interactions by leptin and ghrelin with neural circuits originating in the caudal brainstem also are important for maintenance of energy balance. For example, Ob-Rb mRNA is found in many brainstem nuclei involved in food intake, including the dorsal vagal complex (DVC) (19, 80). We have used similar techniques to demonstrate GHSR expression in many of the same brainstem nuclei (our unpublished observations). The presence of ghrelin and leptin receptors in the brainstem likely contributes to the *c-fos* induction in the brainstem following ghrelin or leptin administration (29, 31, 50, 71, 81). Phosphorylation of STAT-3, which occurs following activation of Ob-Rb, occurs in several brainstem nuclei following peripheral administration of leptin (35). Delivery of leptin to the caudal brainstem reduces food intake and body weight (80). Furthermore, injection of leptin directly into the DVC reduces food intake and body weight, whereas ghrelin injection into the DVC causes hyperphagia [Ref. 80; and Faulconbridge, L. F., and H. J. Grill (University of Pennsylvania, Philadelphia, PA), personal communication].

Downstream Targets of Arcuate Neurons: Effector Arms of the Circuit

The aforementioned data support the model that many of the effects of metabolic cues, such as those of leptin and ghrelin, are mediated by POMC/CART and NPY/AgRP neurons in the Arc. However, the downstream sites that

are targeted by these Arc neurons and that mediate the behavioral, endocrine and autonomic effects of changes in energy status are less understood. Numerous neuro-anatomical and physiologic studies have suggested that leptin-responsive Arc neurons influence the activity of neurons in key effector central sites. These include the paraventricular hypothalamic nucleus (PVH), the lateral hypothalamic area (LHA), the VMH, and autonomic preganglionic neurons in the interomedialateral nucleus of the spinal cord and the DVC (1, 21, 32, 59, 82–84). As outlined in Fig. 1, these putative second-order neurons also contain orexigenic and anorexigenic neuropeptides, such as orexin (hypocretin)-A and -B, cholecystokinin, melanin-concentrating hormone (MCH), and GLP-1. Several of these sites can be linked to the behavioral, hypophysiotropic, and autonomic responses responsible for adapting to changes in energy status. In what follows in this section, we will present briefly three examples of putative circuits through which leptin acts. Each serves as an example of hypothalamic integration of endocrine, autonomic, and behavioral responses.

The TRH Neuroendocrine Response to Fasting

Falls in leptin initiate many neuroendocrine responses to starvation (85). Prominent among these is the inhibition of thyroid hormone secretion, which is thought to be adaptive because of the ensuing drop in the metabolic rate and preservation of energy stores (85, 86). Some examples of leptin's effects on the hypothalamic-pituitary-thyroid axis include blunting of the starvation-induced falls in circulating T₄ levels and TRH mRNA levels in the PVH by administration of leptin to fasted animals (85, 87). This occurs at least in part via projections of leptin-responsive Arc neurons to TRH neurons because monosodium glutamate-induced ablation of the Arc blunts the ability of leptin to prevent fasting-induced falls in TRH gene expression (87–89). More specifically, leptin's effects on the TRH neurons in the PVH are mediated in part through the melanocortin system. Centrally-administered α -MSH or α -MSH analogs can prevent or minimize the fasting-induced suppression of TRH gene expression and drop in thyroid hormone levels, can raise plasma TSH in fasted animals and can increase TRH release from hypothalamic explants (90, 91). Furthermore, central AgRP administration decreases plasma TSH levels in fed animals, produces long-lasting suppression of plasma TSH and circulating T₄ (when injected directly into the PVH), and blocks α -MSH- and leptin-induced TRH release from hypothalamic explants (90). Neuroanatomic support includes the innervation of TRH neurons by α -MSH and AgRP-containing neurons and the coexpression of melanocortin-4 receptor (MC4R) mRNA within TRH-containing PVH neurons (91–93). Leptin also directly interacts with TRH-containing PVH neurons, as evidenced by Ob-Rb mRNA expression within PVH neurons, leptin-induced expression of suppressor of cytokine signaling-3 mRNA in TRH neurons, and leptin activation of the TRH promoter, *in vitro* (19, 92).

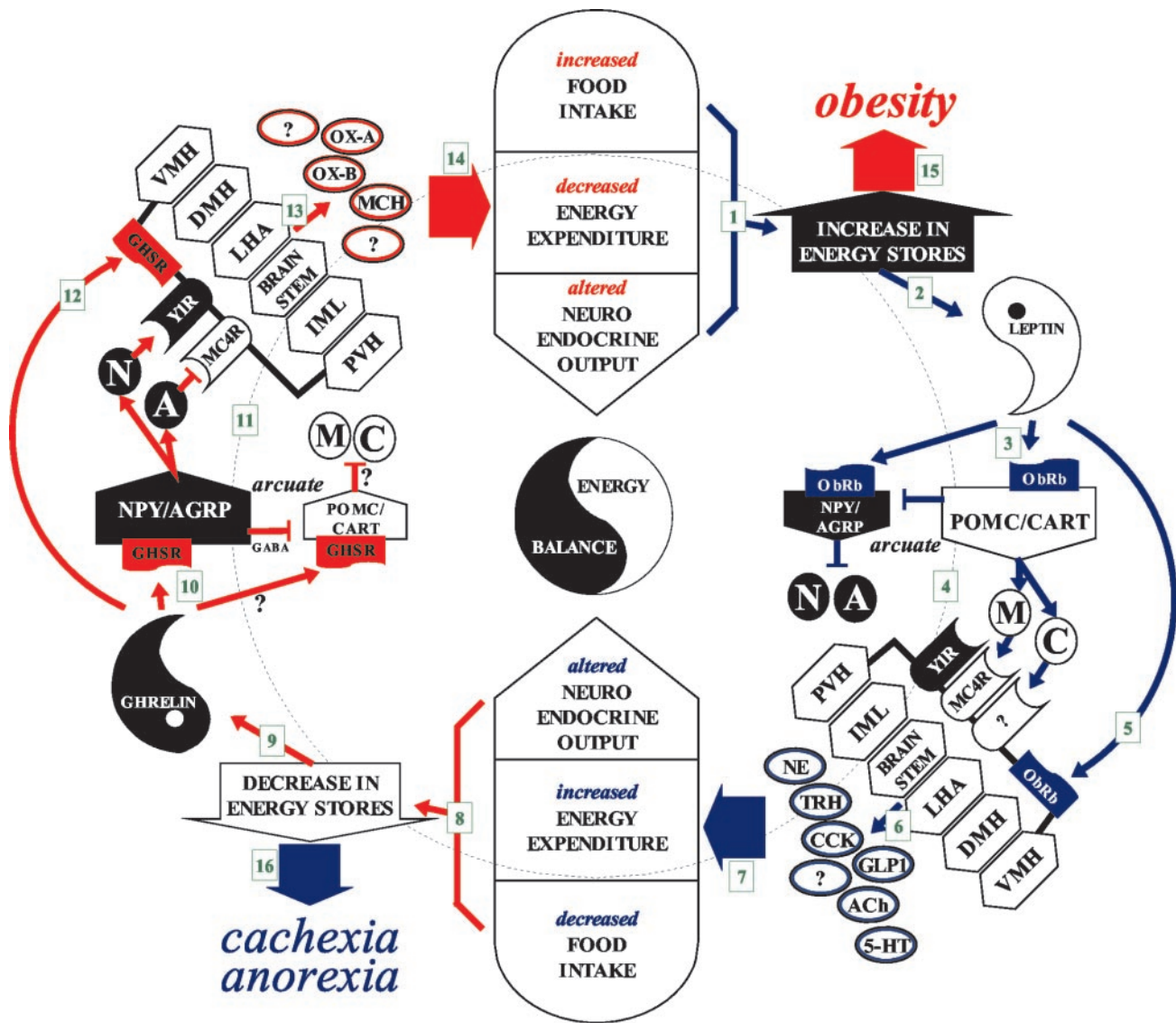


FIG. 1. Pathways involved in energy balance. 1) Increased food intake, decreased energy expenditure and altered neuroendocrine output lead to a relative increase in energy stores. 2) In response, leptin is released from white adipose tissue. 3) Leptin travels to the Arc, where it stimulates POMC/CART neurons and inhibits NPY/AgRP neurons, via interactions with its receptor, Ob-Rb. 4) α -MSH and CART are released at key behavioral, autonomic, and neuroendocrine regulatory regions. 5) These regulatory regions also have direct input from leptin. 6) α -MSH, CART, and leptin influence the release of various neuropeptides and neurotransmitters from these regulatory regions. 7) These substances lead to decreased food intake, increased energy expenditure and altered neuroendocrine output which, in turn, 8) result in a relative decrease in energy stores. 9) In response, ghrelin is released from the GI tract. 10) Ghrelin travels to the Arc, where it activates NPY/AgRP neurons and likely inhibits POMC/CART neurons, via interactions with its receptor, GHSR. 11) NPY and AgRP are released at key behavioral, autonomic, and neuroendocrine regulatory regions. 12) These regulatory regions also have direct input from ghrelin. 13) NPY, AgRP, and ghrelin influence the release of various neuropeptides and neurotransmitters from these regulatory regions. 14) These substances effect responses that lead back to step 1. 15) Dysregulated stimulation of ghrelin-activated pathways and/or blockade of leptin-activated pathways cause obesity. 16) Dysregulated stimulation of leptin-activated pathways and/or blockade of ghrelin-activated pathways cause cachexia.

Leptin, the Autonomic Nervous System, and Energy Expenditure

In an attempt to maintain adequate energy stores, mammals reduce energy expenditure during periods of reduced food intake. Several lines of evidence support a role for reduced sympathetic activity mediating decreased energy expenditure during periods of hypoleptinemia. For example, absence of leptin signaling (*ob/ob* and *db/db* mice) results in decreased energy expenditure and contributes to the obesity in those animals (94). In addition, leptin increases uncou-

pling protein-1 mRNA levels in brown adipose tissue of fasted animals, suggesting that thermogenesis is decreased (95, 96). Furthermore, central leptin administration attenuates the reductions in heart rate, blood pressure, and energy expenditure normally observed during periods of negative energy balance (97–103).

The neuroanatomical substrate for leptin to regulate sympathetic outflow remains to be characterized; however, several studies have suggested a role for the melanocortin system. MC4R antagonists block leptin-stimulated uncoupling

protein-1 gene expression in brown adipose tissue (104). In addition, MC4R mRNA is expressed not only in several CNS sites that provide direct inputs to autonomic preganglionic neurons, but also in those same sympathetic and parasympathetic preganglionic neurons (82, 84). Moreover, leptin-activated arcuate POMC/CART neurons directly innervate sympathetic preganglionic neurons, including the regions of the IML known to innervate brown adipose tissue (32, 105). While the functional significance of this simple circuit remains to be characterized, the aforementioned data suggest that leptin-responsive POMC/CART neurons in the Arc directly target and affect the activity of parasympathetic and sympathetic preganglionic neurons that are critical for regulating coordinated autonomic responses including energy expenditure.

Linking Changes in Leptin and Ghrelin to Ingestive Behavior

Many studies indicate that leptin and its downstream central pathways affect food intake (94, 106–111); see also accompanying review by O'Rahilly *et al.* (37). As discussed above, leptin mediates these effects via leptin action in the CNS. Indeed, selective deletion of neuronal leptin receptors produces obesity (24). Several pieces of evidence suggest that two sites may mediate the effects of leptin to regulate feeding. The first candidate is the PVH, which, as noted above, receives innervation from leptin- and ghrelin-responsive neurons in the Arc (59, 83). The PVH neurons that contribute to food intake modulation include those that innervate autonomic preganglionic neurons and express MC4R mRNA (1, 82, 84). The second candidate population is the LHA. The LHA has been known to play a key role in the regulation of ingestive behavior since early lesion studies (1, 59). In recent years, two peptides were discovered that are expressed in the brain only by neurons in this area: MCH and the orexins (112–114). Notably, the MCH and orexin neurons have widespread projections, including several CNS sites that may be involved in controlling ingestive behavior (1). The receptors for both peptides are similarly widespread and very similar in their distribution (115–117). Central injections of MCH increases food intake in the rat and MCH mRNA levels are increased by food deprivation (118). MCH $-/-$ mice are hypophagic and lean and mice that overexpress MCH are obese and hyperleptinemic (119, 120). The role of orexins in regulating food intake is less well defined, but icv injections of orexin increase feeding behavior, whereas food restriction increases orexin mRNA (113, 121). Interestingly, leptin-responsive POMC/CART and NPY/AgRP neurons innervate MCH and orexin neurons (21, 58, 122). Thus, several pieces of data suggest that MCH and orexin cells in the LHA are downstream of leptin- and ghrelin-responsive neurons in the Arc. These projections may play a key role in regulating feeding behavior during periods of changing energy availability.

Key CNS Pathways Serve as the Battleground for the Control of Energy Balance

As mentioned above, there are many examples whereby dysregulation of leptin- and ghrelin-responsive central pathways result in obesity. In contrast, the CNS circuits underlying

responses at the opposite end of the energy spectrum, and which contribute to cachexia and eating disorders such as anorexia nervosa, are less understood but also involve parallel signaling defects in these same leptin- and ghrelin-responsive circuits. Regarding anorexia nervosa, recent data have tied the actions of the central serotonergic pathway to melanocortin pathways (123). This is relevant in that dysregulation of the central serotonergic system has long been implicated in the pathogenesis of eating disorders such as anorexia nervosa and in that serotonergic agents inhibit food intake and body weight, even in humans (124). Indeed, one of the most effective drugs used to treat human obesity was fenfluramine used in combination with phentermine (Fen/Phen). Although not completely understood, fenfluramine is thought to mediate its effects by increasing serotonin release and inhibiting serotonin reuptake (125). Other evidence supporting a critical role of serotonergic receptors in regulating body weight homeostasis stems from the observation that deletion of serotonin receptors results in obesity. Specifically, deletion of the serotonin 2C receptor (5-HT_{2C}R) induces hyperphagia, obesity, and diabetes (126). Moreover, these mice are insensitive to the anorectic actions of fenfluramine, suggesting that 5-HT_{2C}R is required to mediate the anorectic actions of serotonin (127). However, the neural mechanisms underlying this response have remained unclear.

Recently, Heisler *et al.* (123) found that arcuate POMC neurons express 5-HT_{2C}R mRNA. In addition, threshold doses of fenfluramine to decrease food intake induce *c-fos* expression and directly depolarize POMC neurons. These findings lead to the hypothesis that POMC neurons are downstream mediators of serotonergic pathways to affect food intake and body weight. Supportive of this, genetic or pharmacological blockade of melanocortin receptors blunted the ability of fenfluramine to induce anorexia (123). Linking the anorectic actions of fenfluramine to the melanocortin pathway is a long way from unraveling the complex pathophysiology of eating disorders. Nonetheless, extensions of these findings may shed light on the neuroanatomical substrate underlying anorexia nervosa.

Relatively more evidence has recently accumulated to suggest that the melanocortin pathway is important in mediating the cachectic responses that often accompany chronic infections or neoplastic syndromes (reviewed in Refs. 128 and 129). For example, central administration of AgRP or other melanocortin receptor antagonists suppresses cachexia induced by both lipopolysaccharide (LPS) and tumor growth (49). Moreover, the responses to LPS or chronic tumor load are blunted in MC4R $-/-$ mice (130). In addition, central α -MSH has potent antipyretic effects as it blunts the febrile responses elicited by LPS administration and prevents LPS-induced anorexia in rats (131, 132). However, lesions of the Arc are not sufficient to block IL-1 induced anorexia and may even enhance the response (133). Interestingly, cytokine administration increases the levels of leptin (134, 135).

Ghrelin levels also are elevated in many types of cachexia, including tumor-bearing rats, humans with cardiac cachexia, and humans with anorexia nervosa (49, 136–138). Ghrelin also has greatly reduced orexigenic potency in tumor-bearing rats compared with control animals (49). These findings suggest that, just as leptin resistance is associated with many

forms of obesity, ghrelin resistance may be associated with cachexia. Although inherently difficult to support with experimental data, it is intriguing to speculate that, similar to leptin resistance, ghrelin resistance may have had evolutionary benefit. This apparently paradoxical resistance to ghrelin could prevent sick individuals, before recovery, from spreading illness to others or from transmitting cancer-promoting or disease-promoting genes that result in death to further generations. Obviously, in our current environment of relatively effective infectious disease and cancer treatments, ghrelin resistance may not be of major evolutionary consequence. However, it certainly may have implications in the medical management of patients with cachexia and anorexia.

Summary

Accumulating evidence supports the model that stimulation of leptin- and ghrelin-responsive pathways, including the central melanocortin system, contributes to the maintenance of body weight. As outlined above and as illustrated in Fig. 1, dysregulation of these pathways leads to pathological weight loss that is comprised of both chronic anorexia and inappropriate energy expenditure. On the other end of the energy balance spectrum, dysregulation of these pathways results in a net weight gain and subsequently obesity. Whereas this model is unavoidably oversimplified, it is supported by accumulating evidence that overlapping CNS pathways are operational at either end of the spectrum of energy balance. Moreover, chronic alterations in the system inherently leads to biased responses resulting in pathological weight loss or obesity.

Acknowledgments

Received February 24, 2003. Accepted April 15, 2003.

Address all correspondence and requests for reprints to: Joel K. Elmquist, D.V.M., Ph.D., 325 Research North, 99 Brookline Avenue, Division of Endocrinology, Beth Israel Deaconess Medical Center, Boston, Massachusetts 02215. E-mail: jelmquis@bidmc.harvard.edu.

This work was supported by NIH Grants DK56116, DK53301, MH61583, and 5T32DK07516-18.

References

- Saper CB, Chou TC, Elmquist JK 2002 The need to feed: homeostatic and hedonic control of eating. *Neuron* 36:199–211
- Barsh GS, Schwartz MW 2002 Genetic approaches to studying energy balance: perception and integration. *Nat Rev Genet* 3:589–600
- Schwartz MW, Woods SC, Porte Jr D, Seeley RJ, Baskin DG 2000 Central nervous system control of food intake. *Nature* 404:661–671
- Friedman JM, Halaas JL 1998 Leptin and the regulation of body weight in mammals. *Nature* 395:763–770
- Grill HJ, Kaplan JM 2002 The neuroanatomical axis for control of energy balance. *Front Neuroendocrinol* 23:2–40
- Spiegelman BM, Flier JS 2001 Obesity and the regulation of energy balance. *Cell* 104:531–543
- Batterham RL, Cowley MA, Small CJ, Herzog H, Cohen MA, Dakin CL, Wren AM, Brynes AE, Low MJ, Ghatei MA, Cone RD, Bloom SR 2002 Gut hormone PYY(3–36) physiologically inhibits food intake. *Nature* 418:650–654
- Levin BE, Dunn-Meynell AA, Routh VH 2001 Brain glucosensing and the K(ATP) channel. *Nat Neurosci* 4:459–460
- Flier JS 1998 Clinical review 94: what's in a name? In search of leptin's physiological role. *J Clin Endocrinol Metab* 83:1407–1413
- Friedman JM 2000 Obesity in the new millennium. *Nature* 404:632–634
- Masuzaki H, Paterson J, Shinyama H, Morton NM, Mullins JJ, Seckl JR, Flier JS 2001 A transgenic model of visceral obesity and the metabolic syndrome. *Science* 294:2166–2170
- Ahima RS, Flier JS 2000 Leptin. *Annu Rev Physiol* 62:413–437
- Cusin I, Sainsbury A, Doyle P, Rohner-Jeanrenaud F, Jeanrenaud B 1995 The ob gene and insulin. A relationship leading to clues to the understanding of obesity. *Diabetes* 44:1467–1470
- De Vos P, Saladin R, Auwerx J, Staels B 1995 Induction of ob gene expression by corticosteroids is accompanied by body weight loss and reduced food intake. *J Biol Chem* 270:15958–15961
- Rajala MW, Scherer PE 2003 Minireview: the adipocyte—at the crossroads of energy homeostasis, inflammation, and atherosclerosis. *Endocrinology* 144:3765–3773
- Fei H, Okano HJ, Li C, Lee GH, Zhao C, Darnell R, Friedman JM 1997 Anatomic localization of alternatively spliced leptin receptors (Ob-R) in mouse brain and other tissues. *Proc Natl Acad Sci USA* 94:7001–7005
- Schwartz MW, Seeley RJ, Campfield LA, Burn P, Baskin DG 1996 Identification of targets of leptin action in rat hypothalamus. *J Clin Invest* 98:1101–1106
- Mercer JG, Hoggard N, Williams LM, Lawrence CB, Hannah LT, Trayhurn P 1996 Localization of leptin receptor mRNA and the long form splice variant (Ob-Rb) in mouse hypothalamus and adjacent brain regions by in situ hybridization. *FEBS Lett* 387:113–116
- Elmquist JK, Bjorbaek C, Ahima RS, Flier JS, Saper CB 1998 Distributions of leptin receptor mRNA isoforms in the rat brain. *J Comp Neurol* 395:535–547
- Bjorbaek C, Elmquist JK, Frantz JD, Shoelson SE, Flier JS 1998 Identification of SOCS-3 as a potential mediator of central leptin resistance. *Mol Cell* 1:619–625
- Elias CF, Aschkenasi C, Lee C, Kelly J, Ahima RS, Bjorbaek C, Flier JS, Saper CB, Elmquist JK 1999 Leptin differentially regulates NPY and POMC neurons projecting to the lateral hypothalamic area. *Neuron* 23:775–786
- Cowley MA, Smart JL, Rubinstein M, Cerdan MG, Diano S, Horvath TL, Cone RD, Low MJ, Cerdan MG, Diano S, Horvath TL, Cone RD, Low MJ 2001 Leptin activates anorexigenic POMC neurons through a neural network in the arcuate nucleus. *Nature* 411:480–484
- Spanswick D, Smith MA, Groppi VE, Logan SD, Ashford ML 1997 Leptin inhibits hypothalamic neurons by activation of ATP-sensitive potassium channels. *Nature* 390:521–525
- Cohen P, Zhao C, Cai X, Montez JM, Rohani SC, Feinstein P, Mombaerts P, Friedman JM 2001 Selective deletion of leptin receptor in neurons leads to obesity. *J Clin Invest* 108:1113–1121
- Minokoshi Y, Kim YB, Peroni OD, Fryer LG, Muller C, Carling D, Kahn BB 2002 Leptin stimulates fatty-acid oxidation by activating AMP-activated protein kinase. *Nature* 415:339–343
- Tartaglia LA, Dembski M, Weng X, Deng N, Culpepper J, Devos R, Richards GJ, Campfield LA, Clark FT, Deeds J, Muir C, Sanker S, Moriarty A, Moore KJ, Smutko JS, Mays GG, Wolf EA, Monroe CA, Tepper RI 1995 Identification and expression cloning of a leptin receptor, OB-R. *Cell* 83:1263–1271
- Tartaglia LA 1997 The leptin receptor. *J Biol Chem* 272:6093–6096
- Woods AJ, Stock MJ 1996 Leptin activation in hypothalamus. *Nature* 381:745
- Van Dijk G, Thiele TE, Donahey JC, Campfield LA, Smith FJ, Burn P, Bernstein IL, Woods SC, Seeley RJ 1996 Central infusions of leptin and GLP-1(7–36) amide differentially stimulate c-FLI in the rat brain. *Am J Physiol* 271:R1096–R1100
- Elmquist JK, Ahima RS, Maratos-Flier E, Flier JS, Saper CB 1997 Leptin activates neurons in ventrobasal hypothalamus and brainstem. *Endocrinology* 138:839–842
- Elias CF, Kelly JF, Lee CE, Ahima RS, Drucker DJ, Saper CB, Elmquist JK 2000 Chemical characterization of leptin-activated neurons in the rat brain. *J Comp Neurol* 423:261–281
- Elias CF, Lee C, Kelly J, Aschkenasi C, Ahima RS, Couceyro PR, Kuhar MJ, Saper CB, Elmquist JK 1998 Leptin activates hypothalamic CART neurons projecting to the spinal cord. *Neuron* 21:1375–1385
- Elmquist JK, Ahima RS, Elias CF, Flier JS, Saper CB 1998 Leptin activates distinct projections from the dorsomedial and ventromedial hypothalamic nuclei. *Proc Natl Acad Sci USA* 95:741–746
- Hubschle T, Thom E, Watson A, Roth J, Klaus S, Meyerhof W 2001 Leptin-induced nuclear translocation of STAT3 immunoreactivity in hypothalamic nuclei involved in body weight regulation. *J Neurosci* 21:2413–2424
- Hosoi T, Kawagishi T, Okuma Y, Tanaka J, Nomura Y 2002 Brain stem is a direct target for leptin's action in the central nervous system. *Endocrinology* 143:3498–3504
- Niswender KD, Morton GJ, Stearns WH, Rhodes CJ, Myers Jr MG, Schwartz MW 2001 Intracellular signalling. Key enzyme in leptin-induced anorexia. *Nature* 413:794–795
- O'Rahilly S, Sadaf Farooqi I, Yeo GSH, Challis BG 2003 Minireview: human obesity—lessons from monogenic disorders. *Endocrinology* 144:3757–3764
- Horvath TL, Diano S, Sotonyi P, Heiman M, Tschöp M 2001 Minireview: ghrelin and the regulation of energy balance—a hypothalamic perspective. *Endocrinology* 142:4163–4169
- Kojima M, Hosoda H, Date Y, Nakazato M, Matsuo H, Kangawa K 1999 Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature* 402:656–660
- Howard AD, Feighner SD, Cully DF, Arena JP, Liberato PA, Rosenblum CI, Hamelin M, Hreniuk DL, Palyha OC, Anderson J, Paress PS, Diaz C, Chou M, Liu KK, McKee KK, Pong SS, Chaung LY, Elbrecht A, Dashkevich M, Heavens R, Rigby M, Sirinathsinghji DJ, Dean DC, Melillo DG, Patchett AA, Nargund R, Griffin PR, DeMartino JA, Gupta SK, Schaeffer JM, Smith

- RG, Van der Ploeg LH 1996 A receptor in pituitary and hypothalamus that functions in growth hormone release. *Science* 273:974–977
41. Guan XM, Yu H, Palyha OC, McKee KK, Feighner SD, Sirinathsinghji DJ, Smith RG, Van der Ploeg LH, Howard AD 1997 Distribution of mRNA encoding the growth hormone secretagogue receptor in brain and peripheral tissues. *Brain Res Mol Brain Res* 48:23–29
 42. Gnanapavan S, Kola B, Bustin SA, Morris DG, McGee P, Fairclough P, Bhattacharya S, Carpenter R, Grossman AB, Korbonits M 2002 The tissue distribution of the mRNA of ghrelin and subtypes of its receptor, GHS-R, in humans. *J Clin Endocrinol Metab* 87:2988
 43. Shuto Y, Shibasaki T, Wada K, Parhar I, Kamegai J, Sugihara H, Oikawa S, Wakabayashi I 2001 Generation of polyclonal antiserum against the growth hormone secretagogue receptor (GHS-R): evidence that the GHS-R exists in the hypothalamus, pituitary and stomach of rats. *Life Sci* 68:991–996
 44. Bennett PA, Thomas GB, Howard AD, Feighner SD, van der Ploeg LH, Smith RG, Robinson IC 1997 Hypothalamic growth hormone secretagogue-receptor (GHS-R) expression is regulated by growth hormone in the rat. *Endocrinology* 138:4552–4557
 45. Mitchell V, Bouret S, Beauvillain JC, Schilling A, Perret M, Kordon C, Epelbaum J 2001 Comparative distribution of mRNA encoding the growth hormone secretagogue-receptor (GHS-R) in *Microcebus murinus* (primate, lemurian) and rat forebrain and pituitary. *J Comp Neurol* 429:469–489
 46. Cummings DE, Purnell JQ, Frayo RS, Schmidova K, Wisse BE, Weigle DS 2001 A preprandial rise in plasma ghrelin levels suggests a role in meal initiation in humans. *Diabetes* 50:1714–1719
 47. Cummings DE, Weigle DS, Frayo RS, Breen PA, Ma MK, Dellinger EP, Purnell JQ 2002 Plasma ghrelin levels after diet-induced weight loss or gastric bypass surgery. *N Engl J Med* 346:1623–1630
 48. Tschop M, Smiley DL, Heiman ML 2000 Ghrelin induces adiposity in rodents. *Nature* 407:908–913
 49. Wisse BE, Frayo RS, Schwartz MW, Cummings DE 2001 Reversal of cancer anorexia by blockade of central melanocortin receptors in rats. *Endocrinology* 142:3292–3301
 50. Nakazato M, Murakami N, Date Y, Kojima M, Matsuo H, Kangawa K, Matsukura S 2001 A role for ghrelin in the central regulation of feeding. *Nature* 409:194–198
 51. Asakawa A, Inui A, Kaga T, Yuzuriha H, Nagata T, Ueno N, Makino S, Fujimiya M, Niiijima A, Fujino MA, Kasuga M 2001 Ghrelin is an appetite-stimulatory signal from stomach with structural resemblance to motilin. *Gastroenterology* 120:337–345
 52. Kamegai J, Tamura H, Shimizu T, Ishii S, Sugihara H, Wakabayashi I 2000 Central effect of ghrelin, an endogenous growth hormone secretagogue, on hypothalamic peptide gene expression. *Endocrinology* 141:4797–4800
 53. Wang L, Saint-Pierre DH, Tache Y 2002 Peripheral ghrelin selectively increases Fos expression in neuropeptide Y-synthesizing neurons in mouse hypothalamic arcuate nucleus. *Neurosci Lett* 325:47–51
 54. Tschop M, Weyer C, Tataranni PA, Devanarayan V, Ravussin E, Heiman ML 2001 Circulating ghrelin levels are decreased in human obesity. *Diabetes* 50:707–709
 55. Cummings DE, Clement K, Purnell JQ, Vaisse C, Foster KE, Frayo RS, Schwartz MW, Basdevant A, Weigle DS 2002 Elevated plasma ghrelin levels in Prader Willi syndrome. *Nat Med* 8:643–644
 56. Haq AM, Farooqi IS, O'Rahilly S, Stadler DD, Rosenfeld RG, Pratt KL, LaFranchi SH, Purnell JQ 2003 Serum ghrelin levels are inversely correlated with body mass index, age, and insulin concentrations in normal children and are markedly increased in Prader-Willi syndrome. *J Clin Endocrinol Metab* 88:174–178
 57. Hahn TM, Breininger JF, Baskin DG, Schwartz MW 1998 Coexpression of Agrp and NPY in fasting-activated hypothalamic neurons. *Nat Neurosci* 1:271–272
 58. Broberger C, De Lecea L, Sutcliffe JG, Hokfelt T 1998 Hypocretin/orexin- and melanin-concentrating hormone-expressing cells form distinct populations in the rodent lateral hypothalamus: relationship to the neuropeptide Y and agouti gene-related protein systems. *J Comp Neurol* 402:460–474
 59. Elmquist JK, Elias CF, Saper CB 1999 From lesions to leptin: hypothalamic control of food intake and body weight. *Neuron* 22:221–232
 60. Spiegelman BM, Flier JS 2001 Obesity and the regulation of energy balance. *Cell* 104:531–543
 61. Cheung CC, Clifton DK, Steiner RA 1997 Proopiomelanocortin neurons are direct targets for leptin in the hypothalamus. *Endocrinology* 138:4489–4492
 62. Willesen MG, Kristensen P, Romer J 1999 Co-localization of growth hormone secretagogue receptor and NPY mRNA in the arcuate nucleus of the rat. *Neuroendocrinology* 70:306–316
 63. Schwartz MW, Seeley RJ, Woods SC, Weigle DS, Campfield LA, Burn P, Baskin DG 1997 Leptin increases hypothalamic pro-opiomelanocortin mRNA expression in the rostral arcuate nucleus. *Diabetes* 46:2119–2123
 64. Thornton JE, Cheung CC, Clifton DK, Steiner RA 1997 Regulation of hypothalamic proopiomelanocortin mRNA by leptin in ob/ob mice. *Endocrinology* 138:5063–5066
 65. Mizuno TM, Kleopoulos SP, Bergen HT, Roberts JL, Priest CA, Mobbs CV 1998 Hypothalamic pro-opiomelanocortin mRNA is reduced by fasting and [corrected] in ob/ob and db/db mice, but is stimulated by leptin. *Diabetes [Erratum] (1998) 47:696] 47:294–297*
 66. Ahima RS, Kelly J, Elmquist JK, Flier JS 1999 Distinct physiologic and neuronal responses to decreased leptin and mild hyperleptinemia. *Endocrinology* 140:4923–4931
 67. Mizuno TM, Makimura H, Silverstein J, Roberts JL, Lopingco T, Mobbs CV 1999 Fasting regulates hypothalamic neuropeptide Y, agouti-related peptide, and proopiomelanocortin in diabetic mice independent of changes in leptin or insulin. *Endocrinology* 140:4551–4557
 68. Stephens TW, Basinski M, Bristow PK, Bue-Valleskey JM, Burgett SG, Craft L, Hale J, Hoffmann J, Hsiung HM, Kriauciunas A, Mackellar W, Rosteck PR, Schoner B, Smith D, Tinsley FC, Zhang XY, Heiman M 1995 The role of neuropeptide Y in the antiobesity action of the obese gene product. *Nature* 377:530–532
 69. Tamura H, Kamegai J, Shimizu T, Ishii S, Sugihara H, Oikawa S 2002 Ghrelin stimulates GH but not food intake in arcuate nucleus ablated rats. *Endocrinology* 143:3268–3275
 70. Hewson AK, Dickson SL 2000 Systemic administration of ghrelin induces Fos and Egr-1 proteins in the hypothalamic arcuate nucleus of fasted and fed rats. *J Neuroendocrinol* 12:1047–1049
 71. Lawrence CB, Snape AC, Baudoin FM, Luckman SM 2002 Acute central ghrelin and GH secretagogues induce feeding and activate brain appetite centers. *Endocrinology* 143:155–162
 72. Dickson SL, Luckman SM 1997 Induction of c-fos messenger ribonucleic acid in neuropeptide Y and growth hormone (GH)-releasing factor neurons in the rat arcuate nucleus following systemic injection of the GH secretagogue, GH-releasing peptide-6. *Endocrinology* 138:771–777
 73. Seoane LM, Lopez M, Tovar S, Casanueva FF, Senaris R, Dieguez C 2003 Agouti-related peptide, neuropeptide Y, and somatostatin-producing neurons are targets for ghrelin actions in the rat hypothalamus. *Endocrinology* 144:544–551
 74. Traeber T, Riediger T, Whitebread S, Scharrer E, Schmid HA 2002 Ghrelin acts on leptin-responsive neurons in the rat arcuate nucleus. *J Neuroendocrinol* 14:580–586
 75. Cowley MA, Smith RG, Diano S, Tschop M, Pronchuk N, Grove KL, Strassburger CJ, Bidlingmaier M, Esterman M, Heiman ML, Garcia-Segura LM, Nillni EA, Mendez P, Low MJ, Sotonyi P, Friedman JM, Liu H, Pinto S, Colmers WF, Cone RD, Horvath TL 2003 The distribution and mechanism of action of ghrelin in the CNS demonstrates a novel hypothalamic circuit regulating energy homeostasis. *Neuron* 37:649–661
 76. Hewson AK, Tung LY, Connell DW, Tookman L, Dickson SL 2002 The rat arcuate nucleus integrates peripheral signals provided by leptin, insulin, and a ghrelin mimetic. *Diabetes* 51:3412–3419
 77. Tschop M, Statnick MA, Suter TM, Heiman ML 2002 GH-releasing peptide-2 increases fat mass in mice lacking NPY: indication for a crucial mediating role of hypothalamic agouti-related protein. *Endocrinology* 143:558–568
 78. Horvath TL, Bechmann I, Naftolin F, Kalra SP, Leranth C 1997 Heterogeneity in the neuropeptide Y-containing neurons of the rat arcuate nucleus: GABAergic and non-GABAergic subpopulations. *Brain Res* 756:283–286
 79. Spanswick D, Smith MA, Mirshamsi S, Routh VH, Ashford ML 2000 Insulin activates ATP-sensitive K⁺ channels in hypothalamic neurons of lean, but not obese rats. *Nat Neurosci* 3:757–758
 80. Grill HJ, Schwartz MW, Kaplan JM, Foxhall JS, Breininger J, Baskin DG 2002 Evidence that the caudal brainstem is a target for the inhibitory effect of leptin on food intake. *Endocrinology* 143:239–246
 81. Lee HM, Wang G, Englander EW, Kojima M, Greeley Jr GH 2002 Ghrelin, a new gastrointestinal endocrine peptide that stimulates insulin secretion: enteric distribution, ontogeny, influence of endocrine, and dietary manipulations. *Endocrinology* 143:185–190
 82. Kishi T, Aschkenasi CJ, Lee CE, Mountjoy KG, Saper CB, Elmquist JK 2003 Expression of melanocortin 4 receptor mRNA in the central nervous system of the rat. *J Comp Neurol* 457:213–235
 83. Cowley MA, Pronchuk N, Fan W, Dinulescu DM, Colmers WF, Cone RD 1999 Integration of NPY, AGRP, and melanocortin signals in the hypothalamic paraventricular nucleus: evidence of a cellular basis for the adipostat. *Neuron* 24:155–163
 84. Mountjoy KG, Mortrud MT, Low MJ, Simerly RB, Cone RD 1994 Localization of the melanocortin-4 receptor (MC4-R) in neuroendocrine and autonomic control circuits in the brain. *Mol Endocrinol* 8:1298–1308
 85. Ahima RS, Prabakaran D, Mantzoros C, Qu D, Lowell B, Maratos-Flier E, Flier JS 1996 Role of leptin in the neuroendocrine response to fasting. *Nature* 382:250–252
 86. Flier JS, Harris M, Hollenberg AN 2000 Leptin, nutrition, and the thyroid: the why, the wherefore, and the wiring. *J Clin Invest* 105:859–861
 87. Legrady G, Emerson CH, Ahima RS, Flier JS, Lechan RM 1997 Leptin prevents fasting-induced suppression of prohypothalamic-releasing hormone messenger ribonucleic acid in neurons of the hypothalamic paraventricular nucleus. *Endocrinology* 138:2569–2576
 88. Legrady G, Emerson CH, Ahima RS, Rand WM, Flier JS, Lechan RM 1998 Arcuate nucleus ablation prevents fasting-induced suppression of ProTRH mRNA in the hypothalamic paraventricular nucleus. *Neuroendocrinology* 68:89–97

89. Blake NG, Eckland DJ, Foster OJ, Lightman SL 1991 Inhibition of hypothalamic thyrotropin-releasing hormone messenger ribonucleic acid during food deprivation. *Endocrinology* 129:2714–2718
90. Kim MS, Small CJ, Stanley SA, Morgan DG, Seal LJ, Kong WM, Edwards CM, Abusnana S, Sunter D, Ghatge MA, Bloom SR 2000 The central melanocortin system affects the hypothalamo-pituitary thyroid axis and may mediate the effect of leptin. *J Clin Invest* 105:1005–1011
91. Fekete C, Legradi G, Mihaly E, Tatro JB, Rand WM, Lechan RM 2000 α -Melanocyte stimulating hormone prevents fasting-induced suppression of corticotropin-releasing hormone gene expression in the rat hypothalamic paraventricular nucleus. *Neurosci Lett* 289:152–156
92. Harris M, Aschkenasi C, Elias CF, Chandrankunnel A, Nilni EA, Bjoorbaek C, Elmquist JK, Flier JS, Hollenberg AN 2001 Transcriptional regulation of the thyrotropin-releasing hormone gene by leptin and melanocortin signaling. *J Clin Invest* 107:111–120
93. Sarkar S, Lechan RM 2003 Central administration of neuropeptide Y reduces α -melanocyte-stimulating hormone-induced cyclic adenosine 5'-monophosphate response element binding protein (CREB) phosphorylation in pro-thyrotropin-releasing hormone neurons and increases CREB phosphorylation in corticotropin-releasing hormone neurons in the hypothalamic paraventricular nucleus. *Endocrinology* 144:281–291
94. Pellemounter MA, Cullen MJ, Baker MB, Hecht R, Winters D, Boone T, Collins F 1995 Effects of the obese gene product on body weight regulation in ob/ob mice. *Science* 269:540–543
95. Sivitz WI, Fink BD, Morgan DA, Fox JM, Donohoue PA, Haynes WG 1999 Sympathetic inhibition, leptin, and uncoupling protein subtype expression in normal fasting rats. *Am J Physiol* 277:E668–E677
96. Sivitz WI, Fink BD, Donohoue PA 1999 Fasting and leptin modulate adipose and muscle uncoupling protein: divergent effects between messenger ribonucleic acid and protein expression. *Endocrinology* 140:1511–1519
97. Williams TD, Chambers JB, May OL, Henderson RP, Rashotte ME, Overton JM 2000 Concurrent reductions in blood pressure and metabolic rate during fasting in the unrestrained SHR. *Am J Physiol Regul Integr Comp Physiol* 278:R255–R262
98. Williams TD, Chambers JB, Henderson RP, Rashotte ME, Overton JM 2002 Cardiovascular responses to caloric restriction and thermoneutrality in C57BL/6j mice. *Am J Physiol Regul Integr Comp Physiol* 282:R1459–R1467
99. Swoap SJ 2001 Altered leptin signaling is sufficient, but not required, for hypotension associated with caloric restriction. *Am J Physiol Heart Circ Physiol* 281:H2473–H2479
100. Aizawa-Abe M, Ogawa Y, Masuzaki H, Ebihara K, Satoh N, Iwai H, Matsuoka N, Hayashi T, Hosoda K, Inoue G, Yoshimasa Y, Nakao K 2000 Pathophysiological role of leptin in obesity-related hypertension. *J Clin Invest* 105:1243–1252
101. VanNess JM, Casto RM, Overton JM 1997 Antihypertensive effects of food-intake restriction in aortic coarctation hypertension. *J Hypertens* 15:1253–1262
102. Overton JM, Williams TD, Chambers JB, Rashotte ME 2001 Central leptin infusion attenuates the cardiovascular and metabolic effects of fasting in rats. *Hypertension* 37:663–669
103. Swoap SJ, Boddell P, Baldwin KM 1995 Interaction of hypertension and caloric restriction on cardiac mass and isomyosin expression. *Am J Physiol* 268:R333–R339
104. Satoh N, Ogawa Y, Katsuura G, Numata Y, Masuzaki H, Yoshimasa Y, Nakao K 1998 Satiety effect and sympathetic activation of leptin are mediated by hypothalamic melanocortin system. *Neurosci Lett* 249:107–110
105. Lowell BB, Flier JS 1997 Brown adipose tissue, β 3-adrenergic receptors, and obesity. *Annu Rev Med* 48:307–316
106. Halaas JL, Boozer C, Blair-West J, Fidathusein N, Denton DA, Friedman JM 1997 Physiological response to long-term peripheral and central leptin infusion in lean and obese mice. *Proc Natl Acad Sci USA* 94:8878–8883
107. Halaas JL, Gajiwala KS, Maffei M, Cohen SL, Chait BT, Rabinowitz D, Lallone RL, Burley SK, Friedman JM 1995 Weight-reducing effects of the plasma protein encoded by the obese gene. *Science* 269:543–546
108. Campfield LA, Smith FJ, Guisez Y, Devos R, Burn P 1995 Recombinant mouse OB protein: evidence for a peripheral signal linking adiposity and central neural networks. *Science* 269:546–549
109. Seeley RJ, Yagaloff KA, Fisher SL, Burn P, Thiele TE, van Dijk G, Baskin DG, Schwartz MW 1997 Melanocortin receptors in leptin effects. *Nature* 390:349
110. Farooqi IS, Jebb SA, Langmack G, Lawrence E, Cheetham CH, Prentice AM, Hughes IA, McCamish MA, O'Rahilly S 1999 Effects of recombinant leptin therapy in a child with congenital leptin deficiency. *N Engl J Med* 341:879–884
111. Montague CT, Farooqi IS, Whitehead JP, Soos MA, Rau H, Wareham NJ, Sewter CP, Digby JE, Mohammed SN, Hurst JA, Cheetham CH, Earley AR, Barnett AH, Prins JB, O'Rahilly S 1997 Congenital leptin deficiency is associated with severe early-onset obesity in humans. *Nature* 387:903–908
112. Bittencourt JC, Presse F, Arias C, Peto C, Vaughan J, Nahon JL, Vale W, Sawchenko PE 1992 The melanin-concentrating hormone system of the rat brain: an immuno- and hybridization histochemical characterization. *J Comp Neurol* 319:218–245
113. Sakurai T, Amemiya A, Ishii M, Matsuzaki I, Chemelli RM, Tanaka H, Williams SC, Richardson JA, Kozlowski GP, Wilson S, Arch JR, Buckingham RE, Haynes AC, Carr SA, Annan RS, McNulty DE, Liu WS, Terrett JA, Elshourbagy NA, Bergsma DJ, Yanagisawa M 1998 Orexins and orexin receptors: a family of hypothalamic neuropeptides and G protein-coupled receptors that regulate feeding behavior. *Cell* 92:573–585
114. de Lecea L, Kilduff TS, Peyron C, Gao X, Foye PE, Danielson PE, Fukuhara C, Battenberg EL, Gautvik VT, Bartlett 2nd FS, Frankel WN, van den Pol AN, Bloom FE, Gautvik KM, Sutcliffe JG 1998 The hypocretins: Hypothalamus-specific peptides with neuroexcitatory activity. *Proc Natl Acad Sci USA* 95:322–327
115. Marcus JN, Aschkenasi CJ, Lee CE, Chemelli RM, Saper CB, Yanagisawa M, Elmquist JK 2001 Differential expression of orexin receptors 1 and 2 in the rat brain. *J Comp Neurol* 435:6–25
116. Kilduff TS, de Lecea L 2001 Mapping of the mRNAs for the hypocretin/orexin and melanin-concentrating hormone receptors: networks of overlapping peptide systems. *J Comp Neurol* 435:1–5
117. Saito Y, Cheng M, Leslie FM, Civelli O 2001 Expression of the melanin-concentrating hormone (MCH) receptor mRNA in the rat brain. *J Comp Neurol* 435:26–40
118. Qu D, Ludwig DS, Gammeltoft S, Piper M, Pellemounter MA, Cullen MJ, Mathes WF, Przypek R, Kanarek R, Maratos-Flier E 1996 A role for melanin-concentrating hormone in the central regulation of feeding behaviour. *Nature* 380:243–247
119. Shimada M, Tritos NA, Lowell BB, Flier JS, Maratos-Flier E 1998 Mice lacking melanin-concentrating hormone are hypophagic and lean. *Nature* 396:670–674
120. Ludwig DS, Tritos NA, Mastaitis JW, Kulkarni R, Kokkotou E, Elmquist J, Lowell B, Flier JS, Maratos-Flier E 2001 Melanin-concentrating hormone overexpression in transgenic mice leads to obesity and insulin resistance. *J Clin Invest* 107:379–386
121. Clegg DJ, Air EL, Woods SC, Seeley RJ 2002 Eating elicited by orexin-a, but not melanin-concentrating hormone, is opioid mediated. *Endocrinology* 143:2995–3000
122. Elias CF, Saper CB, Maratos-Flier E, Tritos NA, Lee C, Kelly J, Tatro JB, Hoffmann GE, Ollmann MM, Barsh GS, Sakurai T, Yanagisawa M, Elmquist JK 1998 Chemically defined projections linking the mediobasal hypothalamus and the lateral hypothalamic area. *J Comp Neurol* 402:442–459
123. Heisler LK, Cowley MA, Tecott LH, Fan W, Low MJ, Smart JL, Rubinstein M, Tatro JB, Marcus JN, Holstege H, Lee CE, Cone RD, Elmquist JK 2002 Activation of central melanocortin pathways by fenfluramine. *Science* 297:609–611
124. Kaye WH, Gwirtsman HE, George DT, Ebert MH 1991 Altered serotonin activity in anorexia nervosa after long-term weight restoration. Does elevated cerebrospinal fluid 5-hydroxyindoleacetic acid level correlate with rigid and obsessive behavior? *Arch Gen Psychiatry* 48:556–562
125. Weintraub M 1992 Long-term weight control: the National Heart, Lung, and Blood Institute funded multimodal intervention study. *Clin Pharmacol Ther* 51:581–585
126. Nonogaki K, Strack AM, Dallman MF, Tecott LH 1998 Leptin-independent hyperphagia and type 2 diabetes in mice with a mutated serotonin 5-HT_{2C} receptor gene. *Nat Med* 4:1152–1156
127. Vickers SP, Clifton PG, Dourish CT, Tecott LH 1999 Reduced satiating effect of d-fenfluramine in serotonin 5-HT_{2C} receptor mutant mice. *Psychopharmacology (Berl)* 143:309–314
128. Marks DL, Cone RD 2001 Central melanocortins and the regulation of weight during acute and chronic disease. *Recent Prog Horm Res* 56:359–375
129. Lechan RM, Tatro JB 2001 Hypothalamic melanocortin signaling in cachexia. *Endocrinology* 142:3288–3291
130. Marks DL, Ling N, Cone RD 2001 Role of the central melanocortin system in cachexia. *Cancer Res* 61:1432–1438
131. Huang QH, Entwistle ML, Alvaro JD, Duman RS, Hruby VJ, Tatro JB 1997 Antipyretic role of endogenous melanocortins mediated by central melanocortin receptors during endotoxin-induced fever. *J Neurosci* 17:3343–3351
132. Huang QH, Hruby VJ, Tatro JB 1999 Role of central melanocortins in endotoxin-induced anorexia. *Am J Physiol* 276:R864–R871
133. Reyes TM, Sawchenko PE 2002 Involvement of the arcuate nucleus of the hypothalamus in interleukin-1-induced anorexia. *J Neurosci* 22:5091–5099
134. Finck BN, Johnson RW 1999 Intracerebroventricular injection of lipopolysaccharide increases plasma leptin levels. *Neuroreport* 10:153–156
135. Grunfeld C, Zhao C, Fuller J, Pollack A, Moser A, Friedman J, Feingold KR 1996 Endotoxin and cytokines induce expression of leptin, the ob gene product, in hamsters. *J Clin Invest* 97:2152–2157
136. Nagaya N, Uematsu M, Kojima M, Date Y, Nakazato M, Okumura H, Hosoda H, Shimizu W, Yamagishi M, Oya H, Koh H, Yutani C, Kangawa K 2001 Elevated circulating level of ghrelin in cachexia associated with chronic heart failure: relationships between ghrelin and anabolic/catabolic factors. *Circulation* 104:2034–2038
137. Tolle V, Kadem M, Bluet-Pajot MT, Frere D, Foulon C, Bossu C, Dardennes R, Mounier C, Zizzari P, Lang F, Epelbaum J, Estour B 2003 Balance in ghrelin and leptin plasma levels in anorexia nervosa patients and constitutionally thin women. *J Clin Endocrinol Metab* 88:109–116
138. Otto B, Cuntz U, Fruehauf E, Wawarta R, Folwaczny C, Riepl RL, Heiman ML, Lehnert P, Fichter M, Tschop M 2001 Weight gain decreases elevated plasma ghrelin concentrations of patients with anorexia nervosa. *Eur J Endocrinol* 145:669–673